# **Identification and cholesterol quantification of tow density lipoprotein subclasses in young adults by VAP-I** I **methodology**

**Krishnaji R Kullrami, David W. Garber, Martin K. Jones, and Jere P. Segrest'** 

Departments of Medicine and Biochemistry and The Atherosclerosis Research Unit, The University of Alabama at Birmingham, Birmingham, AL **35294** 

**Abstract** Low density lipoprotein **(LDL)** particles are heterogeneous in size, density, and chemical composition; small, dense LDL may be more atherogenic than large, buoyant LDL. We have developed a rapid microscale method called LDL VAP-II (Vertical Auto Profile-II) for quantification of cholesterol in LDL subclasses. The method is based upon a short **(1** h) single vertical spin density-gradient ultracentrifugation and on-line VAP-I1 analyzer. LDL VAP-I1 is rapid and reproducible. Using this method five LDL subclasses, designated **as** LDLl (most buoyant) through LDL5 (most dense), have been identified in a population consisting of **195**  medical students (ages, **22-29** years). The R/(relative position of the major LDL peak in the density gradient; the higher the  $R_i$  value, the lower the peak density) was significantly positively correlated with cholesterol levels of high density lipoprotein (HDL) *(7* = **0.594),** HDL3 **(0.350)** and HDLz **(0.625),**  and significantly negatively correlated with triglycerides (TG) **(-0.355)** and cholesterol levels of very low density lipoprotein (VLDL) **(-0.386)** and intermediate density lipoprotein (IDL) **(-0.432).** These results are consistent with those obtained by other investigators. The *R/* value was significantly correlated with peak particle diameter as determined by non-denaturing gradient gel electrophoresis  $(r = 0.859)$ . In a forward stepwise multivariate analysis comparing  $R_f$  with sex, VLDL, LDL, Lp[a], IDL, HDL<sub>3</sub>, HDL<sub>2</sub>, and triglyceride, only HDL<sub>2</sub> remained in the model.-Kulkami, **K. R., D. W. Garber, M. K. Jones, and J. P. Segrest.** Identification and cholesterol quantification of low density lipoprotein subclasses in young adults by VAP-II methodology. *J. Lipid Res.* 1995. 36: **229 1-2302.** 

**Supplementary key words** LDL heterogeneity  $\bullet$  single vertical spin **densitygradient ultracentrifugation gradient gel electrophoresis** 

Elevated plasma concentrations of low density lipo-<br> **Abbreviations: HDL, high density lipoproteins; LDL, low density** protein (LDL) cholesterol are strongly and positively linked to increased risk of coronary heart disease (CHD) (1). **LDL** particles are heterogeneous in their physical and chemical properties, including hydrated density, particle size and shape, and chemical composition. Several subclasses of LDL with differing physicochemical

properties are known to exist in patients with hypertriglyceridemia *(2),* **as** well **as** in normal subjects (3).

There has been an increasing interest in LDL subclasses recently due to the finding from several casecontrol studies that small, dense LDL is more prevalent in CHD patients than in control subjects **(4-7),** thus linking small, dense LDL to increased risk **of** CHD. The increased presence of small, dense LDL is also associated with insulin resistance states such **as** are seen in patients with non-insulindependent diabetes mellitus *(8),* abdominal obesity (9) and hypertension (9, 10). Using non-denaturing gradient gel electrophoresis, Krauss and Burke (11) have identified seven LDL size **sub**classes, and Austin et al. (12) observed two distinct patterns of distribution of these subclasses in human plasma corresponding to two phenotypes: pattern A, comprising mainly large, buoyant LDL particles with peak particle diameter *>255* A, and pattern B, with the predominance of small, dense LDL particles having peak particle diameter *5255* A. In a case-control study (13), a 3-fold increase in CHD risk was attributed to the presence of LDL pattern B, although the significance of its association with CHD disappeared when contributions due **to** triglycerides (TG) and high density lipopre tein (HDL) cholesterol were taken into consideration. However, the presence of pattern B has been shown to be strongly associated with the high risk lipoprotein

lipoproteins; IDL, intermediate density lipoproteins; VLDL, very low **TC, total cholesterol; TG, triglycerides; VAP, Vertical Auto Profile; density lipoproteins; Lp[a], lipoprotein[a]; apo[a], apolipoprotein[a];**  Disease Control and Prevention; NHLBI, National Heart, Lung, and **Blood Institute; NCEP, National Cholesterol Education Program.** 

**<sup>&#</sup>x27;To whom correspondence should be addressed at: DREB Box 3. Room D-630, Birmingham, AL 352940012.** 

profile characterized by increased levels of TG, apolipoprotein (apo) B, intermediate density lipoprotein (IDL), very low density lipoprotein (VLDL), and decreased levels of HDL cholesterol and apoA-I **(12).** Thus, pattern B (small, dense LDL) may serve as a marker of an atherogenic lipoprotein profile.

Recent reports have also suggested that the LDL size distribution is, in part, genetically controlled and that the LDL subclass pattern B may be linked to multiple genes, including the LDL receptor locus on the short arm of chromosome **19p,** the apoA-I/C-III/A-IV cluster on chromosome **16,** and the manganese superoxide dismutase gene on chromosome 6 **(4, 14).** However, environmental effects are known to influence the LDL size distribution as well **(15).** The expression of LDL subclass pattern B is also found to be agedependent, usually expressed only after the age of **20** in men and after menopause in women.

Among various techniques available for LDL subclass analysis, non-denaturing gradient gel electrophoresis and ultracentrifugation methods, including sequential flotation **(16),** analytical **(17),** and various density-gradient methods **(18-21),** have been widely used. All currently used ultracentrifugation procedures are lengthy and laborious, and thus are restricted to research studies. Nondenaturing **2-16%** polyacrylamide gradient gel electrophoresis is the most common method **(1 1,15,22, 23)** because it requires only a small amount of plasma, can analyze multiple samples on a single gel, and can be performed in most laboratories. However, the lack of availability of commercial gels could make reproducibility a problem for new users of this technique. Thus,

 $0.5$ 

**HDL** +Lp{a}

additional methods will be beneficial for the applications involving LDL subclasses.

Here we describe a rapid and reproducible method called LDL VAP-I1 (Vertical Auto Profile-11) which requires less than **3** h for the complete analysis of eight samples while using only 70  $\mu$ l of plasma per sample. Clinical application of this technique to the study of LDL subclass distribution in a healthy population consisting of **195** medical students is demonstrated.

## MATERIALS AND METHODS

#### **Study subjects**

Blood samples from **195** fasting medical students from three different classes **(68** women and **127** men, ages **22-29** years) attending the University of Alabama at Birmingham were drawn into EDTAcontaining Vacutainer tubes, and plasma was separated from the blood by low speed centrifugation. As part of education and cholesterol awareness, each year sophomore medical students are provided with their complete lipoprotein cholesterol profile test performed by the Atherosclerosis Research Unit at the University of Alabama at Birmingham using the VAP-I method described previously **(24-26).** Aliquots from these plasma samples were used for LDL VAP-I1 analysis and gradient gel electrophoresis.

## Lipoprotein **analysis**

dense LDL

 $0.4$ 

Total and lipoprotein [HDL, HDL<sub>3</sub>, HDL<sub>2</sub>, lipoprotein $[a]$  (Lp $[a]$ ), LDL, IDL, and VLDL] cholesterol con-

B

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A

**Fig. 1. Representative examples of** LDL **VAP-I1 absorbance curves (profiles) as recorded by the chart recorder. LDL subclasses were separated by single vertical spin density-gradient ultracentrifugation, and cholesterol distribution in the gradient was measured using the on-line continuous flow VAP-I1 analyzer to obtain absorbance curves corresponding to** LDL **VAP-11 profiles.** 

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**Fig. 2.** Frequency distribution diagram showing the distribution of  $R_f$  (relative position of the major LDL peak **maximum in LDL VAP-I1 gradient) in 195 medical students into five clusters corresponding to five LDL**  subclasses.  $R_f$  of the major LDL peak (LDL peak with the highest peak height) for each subject was calculated **as the ratio of the distance of major LDL peak maximum from the beginning of HDL** + **Lp[a] peak to the distance of IDL + VLDL peak maximum from the beginning of HDL + Lp[a] peak. The mid-R<sub>f</sub> of each cluster is indicated by an arrow.** 

centrations were determined using the VAP-I method (24-26). Triglyceride was measured enzymatically (GPO-Trinder, Sigma Chemical Co., St. Louis, MO) in a subset consisting of 102 medical students.

# **LDL VAP-Il method**

LDL subclass distribution was determined using the LDL VAP-I1 method. Methodology is similar to the VAP-I1 method (27,28), except that the density gradient is adjusted to optimally separate LDL subclasses. In this method, LDL subclasses are first separated by a single vertical spin density gradient ultracentrifugation technique using the whole plasma, and then the cholesterol concentration of each subclass is determined by measuring the cholesterol distribution across the gradient using the VAP-I1 on-line cholesterol analyzer. As described previously, loss of resolution due to mixing of lipoprotein peaks during analysis is minimal in the VAP-I1 cholesterol analyzer compared to the VAP-I analyzer.

Plasma samples (70  $\mu$ l) were diluted 10-fold with 1.006 **g/ml** saline solution prior to the density adjustment because of the high sensitivity of the VAP-I1 analyzer. Density of the diluted plasma was adjusted to 1.08 g/ml **as** follows: 177 **pl** of 1.28 g/ml saline solution was pipetted into a glass tube and dried in a Speed Vac (Virus Co., Inc., Gardiner, NY) for 2 h with heat. Diluted plasma (650 **pl)** was added to the glass tube which was

then vortexed. Density gradients were formed in Quick-Seal polyallomer tubes (Beckman Instruments, Cat. No. 342412) by first pipetting 4.65 ml of 1.041  $g$ /ml saline into a glass Pasteur pipette placed in the polyallomer tube and then underlayering the diluted plasma by pipetting 650 pl of the density-adjusted plasma into the Pasteur pipette. The densities of the plasma and the saline solution used to form the gradient were chosen so **as** to spread the distribution of LDL subclasses along much of the gradient while compressing HDL and Lp[a] towards the bottom and IDL and VLDL towards the top of the gradient. The tubes were sealed and placed immediately in a VTi-80 rotor and centrifuged in a L8-80M model ultracentrifuge (both of Beckman Instruments) using the following conditions: speed, 80,000 rpm;  $\mathbf{\bar{\omega}}^2 t$ ,  $1.80 \times 10^{11}$ ; temperature,  $20^{\circ}$ C; acceleration and deceleration settings, 6. Total time of centrifugation including deceleration was 1 h. After centrifugation, cholesterol distribution across the gradient was measured by continuously drawing the gradient from the bottom of the tube into the VAP-I1 on-line cholesterol analyzer. (Note that the gradient is not collected in test tubes for individual determinations; instead a continuous analysis is performed on the entire gradient to obtain a continuous absorbance curve; for details see references 27 and *28.)* The same optimum analysis conditions described previously for the measurement of major lipoproteins (27) were used.



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*0.5* 

**D** LDL-4 LDI **0** *50* **loo 150 200 250 Gradient Pwltion (RoLIWe)**  LDL-5 LDL-5 Absorbance  $\mathbf{0}$ **0 50 100 150 200 250 Gradient Position (Relative)** 

**LDL-3** 

**LDL-1** 

**LDL-1** 

150

nt Position (Relative)

**LDL-2** 

פ- וחוג

200

100

250

1 **AC LDL-3** 

**Fig. 3.** Computer deconvoluted LDL VAP-II profiles illustrating *matters* (25). LDL subclasses Cholesterol concentrations of all LDL sub previously described (12). **classes in each profile are calculated by deconvoluting the digitized output of LDL VAP-I1 profile as described in Materials and Methods using software developed in this laboratory. Each profile shows the major LDL subclass and minor LDL subclasses that are present. LDL subclasses are shown by solid curves and the parent absorbance curve (the absorbance curve prior to deconvolution) is shown by empty circles (each circle representing a data point) connected by solid line.** 

The absorbance curve corresponding to the cholesterol distribution in the centrifuge tube was monitored by a strip chart recorder, while digitized absorbance data were collected using an analog-to-digital conversion board and a personal computer. The data points are collected at the rate of 60 points per min. As it requires approximately *6.5* min for the complete analysis of me sample, approximately **400** data points are stored for each profile. However, the number of data points for all profiles is normalized to *a* predetermined value (usually the data point number for IDL + VLDL peak maximum being adjusted to **200)** in order to be able to compare profiles with slightly varying analysis times. A tube containing a plasma sample whose total cholesterol concentration was measured by the Northwest Lipid Research Laboratories (NWLRL) at Seattle, WA was included in each rotor in order to convert the area under each LDL VAP-I1 absorbance curve into its cholesterol concentration. The digitized and normalized absorbance curve was mathematically deconvoluted into its components (subcurves) using software developed in this laboratory, providing cholesterol values of *six* LDL subclasses. The software was developed by defining a set of exponential gaussian subcurves for each lipoprotein class. The numbers of subcurves defined for the lipoprotein classes were: LDL, 6; HDL + Lp[a], **3;** and IDL + VLDL, 3. The rationale for defining six subcurves for LDL is described in the Results section. The shape parameters (widths at half height and the exponential parameter) of subcurves of all lipoprotein classes were adjusted to provide optimal fit of the lipoprotein class under most conditions. During a profile deconvolution, peak heights for the pre-defined subcurves for all lipoprotein classes were simultaneously varied until the sum of the squared deviations between the sum of the subcurves and the parent absorbance curve was minimized using a linear regression method.

# **Nondenaturing polyacrylamide gradient gel electrophoresis**

Non-denaturing **2-** 16% polyacrylamide gradient gel electrophoresis was performed on plasma samples obtained from a subset consisting of **32** students at the Donner Laboratory, University of California at Berkeley, CA, as described previously **(29).** Gels were stained for lipid with 0.1 M Oil Red 0, and LDL peak particle diameter was determined using calibration markers **(29).** LDL subclass patterns were assigned **as** 

#### **Statistical**

Cholesterol and triglyceride values are expressed **as**  mean  $(mg/dl) \pm SD$ . Tests of significance were performed using Student's  $t$  test (paired or unpaired as



**Fig. 4. LDL VAP-I1 absorbance curves (profiles) demonstrating within-rotor reproducibility of the method. Eight aliquots of the same plasma sample were centrifuged within a single rotor using the conditions described for LDL VAP-I1 in Materials and Methods. Absorbance curves were obtained by continuous analysis of the contents of the centrifuged tubes for cholesterol using VAP-I1 analyzer.** 

required). Simple linear regression was used to calculate Pearson correlation coefficients for the comparison of two measurements or two techniques. Stepwise multivariate regression analysis was used to compare the relative association of various lipoprotein cholesterols, TG, and sex with LDL  $R_f$ (relative position of the major LDL peak in the density gradient).

## RESULTS

#### **LDL VAP-II and its characterization**

Figure **1** shows four representative examples (A-D) of LDL VAP-I1 cholesterol absorbance curves (profiles) **as** recorded on the chart paper. Two peaks of LDL, corresponding to dense and buoyant LDL, are apparent in all profiles except profile D; thus, LDL VAP-I1 is able to differentiate at least two LDL peaks in most cases. Furthermore, profile A is characterized by the predominance of buoyant LDL, whereas profile B is characterized by the predominance of dense LDL. The presence of nearly equal proportions of both the dense and buoyant LDL peaks can occasionally be observed as shown by profile C, while a single broad LDL peak with an intermediate density is seen in profile D.

Although only two LDL peaks are observed in most LDL VAP-I1 profiles, these peaks do not always occur at the same density positions. LDL subclasses distribute around several density positions, **as** determined by the  $R_f$  distribution. ( $R_f$  is the relative position of the major LDL peak maximum in the density gradient and is calculated by dividing the distance of LDL peak maximum measured from the beginning of the  $HDL + Lp[a]$ peak by the distance of IDL + VLDL peak maximum, also measured from the beginning of the HDL + Lp[a] peak.) **Figure 2** shows the frequency distribution of  $R_f$ values of the major LDL peak in LDL VAP-I1 profiles of all 195 medical students. The figure indicates that the  $R_f$  values distribute into five clusters in this population, suggesting the existence of at least five LDL subclasses, each cluster corresponding to one LDL subclass. The LDL subclasses are designated **as** LDL-1 (most buoyant) through LDL-5 (most dense). The underlying reason for relating one cluster to one subclass is the assumption that all subjects with **the** same major LDL subclass will have similar  $R_f$  values, varying slightly from one another due to the experimental variation, and therefore form a cluster around a mid- $R_f$  value. The  $R_f$  values corresponding to the mid-positions of these five clusters were, in turn, used to assign peak maximal positions for the five LDL subcurves in LDL VAP-I1 profile. However, in several profiles a minor but distinct peak (similar to the dense LDL peak seen in profile A of Fig. **l),** even denser than LDL5, was noticed. Therefore, a sixth subcurve (LDL-6) with its peak position corresponding to the position of this minor peak in the main profile was included.

Mathematical deconvolution software was developed utilizing these LDL subclass peak positions and other peak parameters **as** described in Materials and Methods to determine cholesterol concentrations of LDL sub-

classes. **Figure 3** shows examples of deconvoluted **LDL**  VAP-I1 profiles displaying both the major **LDL** subclass and minor **LDL** subclasses present. The major subclass curve in each example has a corresponding peak in the main profile (absorbance curve). The predominant **LDL**  subclass (the subclass with the highest cholesterol concentration) in each profile was used to classify the profile, thus **LDLl, LDL-2, LDL3, LDL-4,** and **LDL-5**  are the major subclasses in profiles A, B, C, **D,** and E, respectively. The cholesterol concentrations of the subclasses are determined by measuring the area under each subcurve and converting it into cholesterol concentration using a calibrator plasma of known cholesterol content. The software also provides a printout with cholesterol values assigned to each of the six **LDL** subclasses (including the minor **LDL-6** subclass) and graphical output of the deconvoluted profile. Absence of any minor subclass results in a subcurve with negative area and thus a less than optimal curve fit. Therefore, subcurves with negative areas are sequentially excluded from the deconvolution process until an optimal fit is obtained. A zero value is assigned to each excluded subclass and thus is not displayed in the graphical output.

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Reproducibility of the **LDL** VAP-I1 method was **as**sessed using **28** aliquots of the same plasma sample in four rotors, each rotor containing 7 aliquots in addition to an aliquot of a calibration plasma. Within-rotor reproducibility values of the assay, calculated as the coefficient of variation (CV) of the cholesterol concentration of the major **LDL** subclass, for the four rotors were: **1.9%, 8.396, 3.6%,** and **4.6%,** respectively. Excellent within-rotor reproducibility of the method is also demonstrated in **Fig. 4** by the superimposable **LDL** VAP-I1 cholesterol absorbance curves obtained from 8 aliquots of the same sample. Between-rotor CV calculated for the above four rotors was **5.2%.** Long-term reproducibility **of** this method studied using **13** aliquots of a single sample in **13** rotors over a period of several days also yielded a highly satisfactory CV of **4.6%.** The high reproducibility of this method is due to the continuous nature of the analysis. This considerably reduces the variability usually observed when multiple determinations are made on fractions collected in test tubes.

In order to assess the relationship between the density (as measured by the  $R_f$ ) and the size of the major LDL peak, **2- 16%** non-denaturing polyacrylamide gradient gel electrophoresis analyses were performed on a subset consisting of **32** samples at the Donner Laboratory, University of California, Berkeley, CA. *Rf* of the major **LDL** peak in **LDL** VAP-I1 profile significantly correlated with the peak particle diameter of the predominant **LDL**  peak present in the corresponding densitometric scan of the gradient gel electrophoresis. **(Fig. 5;**  $r = 0.859$ ;  $P$ 

< **0.0001).** In addition, each subject was assigned an **LDL**  size distribution pattern **as** A, B, or AB (intermediate pattern) by the Donner Laboratory. Of the **32** subjects, **59%** had pattern A, **34%** pattern AB, and **6%** pattern B. The same **32** subjects were also classified based on their major **LDL** subclass as obtained from **LDL** VAP-11. Comparison of the two classifications suggested that among all pattern A subjects, 68% had either **LDL-1** or **LDLS; 26%, LDL3; 5%, LDL-4;** and 0%, **LDL5 as** their major **LDL** subclass. Among the subjects with pattern AB, **55%** had **LDL4.27% LDL3,** and **18% LDL5 as** their major **LDL** subclass. All subjects with pattern B had **LDL-5 as** their major **LDL** subclass. Thus, **LDLl** and **LDL-2** can be considered to be as equivalent to pattern A, **LDL-3** and **LDL4 as** pattern AB, and **LDL5** as pattern **B.** It is also noteworthy that subjects with **LDL5** who were classified **as** having pattern AB by the gel analysis also showed an additional peak with nearly equal peak height and area as that of the predominant **LDL** peak with its peak particle diameter **<255** Å, further suggesting **LDL5** to be more similar to pattern B **LDL.** These data suggest a reasonable agreement between the two classifications that are based on two different physical properties. [I](http://www.jlr.org/). In addition, each subject was assigned an LDL<br>
ibution pattern as A, B, or AB (intermediate<br>
by the Donner Laboratory, Of the 22 subjects.<br>
pattern A, 54% pattern A6.<br>
22 subjects were also classified based on their<br>



**Fig. 5.** Comparison of  $R_f$  of the major LDL peak in LDL VAP-II **profile with the peak particle diameter of the major LDL peak obtained from the gradient gel electrophoresis. Plasma aliquots ob tained from a subset consisting of 32 samples were used for the analysis by LDL VAP-I1 and by 2-16% nondenaturing gradient gel electrophoresis. Gradient gel electrophoresis analysis was performed at Donner Laboratory, University of California, Berkeley.** *R/* **is the relative peak position of the major LDL peak maximum in the density**  gradient; higher  $R_f$  indicates lower peak density.

#### *Clinical* **application of LDL VAP-11**

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The mean concentration values of total cholesterol, triglyceride,  $R_f$  of the major LDL peak, and various lipoprotein cholesterol fractions obtained for women, men, and for **all** subjects together, are presented in **Table 1.** *As* the *VAP* method separates both Lp[a] and IDL from the "real LDL" (LDLR), these two lipoprotein measurements were also included in the present study. However, in order to be **also** consistent with LDL values reported by others, including NCEP, Lp[a] and IDL cholesterol values were added to the LDLR and the sum is referred to **as** LDL(NCEP). The mean values of all lipoproteins for both women and men were typical. HDL,  $HDL<sub>3</sub>$ , and  $HDL<sub>2</sub>$  cholesterol values in women were significantly higher than in men, while IDL and VLDL cholesterol were significantly higher in men than in women. Although TG values were higher in men than women, the difference in the mean values did not reach significance  $(P > 0.05)$ . Total, Lp[a], LDL-R, and LDL(NCEP) cholesterol did not significantly differ in the two groups. The mean  $R_f$  of the major LDL peak for women was significantly higher than men ( $P \le 0.001$ ) suggesting that women generally had more buoyant LDL than men. The frequency distribution diagram of  $R_f$  of the major LDL peak for women and men (Fig.  $6$ ) also suggested that women tend to have more buoyant LDL than men. This is apparent by the skewing of the  $R_f$  distribution somewhat towards higher  $R_f$  values for women. On the contrary, distribution was skewed towards lower  $R_f$  in the case of men. The  $R_f$  value for the 50th percentile distribution for women was 0.505 compared to 0.465 found for men ( $P \le 0.0001$ ). In addition, LDL subclasses in women appeared to be less dense compared **to** their respective subclasses in men, **as** indicated by a shift in the positions of the clusters towards the right in the diagram shown for women. (The respective mid- $R_f$  values for clusters corresponding to LDL-1, -2, -3, **4,** and -5 for women are **0.62,** 0.555, 0.505, 0.45, and 0.38, while the corresponding values for men are 0.565, 0.515,0.47, 0.415, and 0.34, respectively.)

The association of  $R_f$  of the major LDL peak with various lipoproteins **was** determined from the Pearson product-moment correlation coefficients calculated from  $R_f$  and the respective lipoprotein values. The results are shown in **Table 2.**  $R_f$  correlated strongly positively with HDL, HDL<sub>3</sub>, and HDL<sub>2</sub> cholesterol, and negatively with IDL and VLDL cholesterol, and with TG when **all** subjects were considered together. Although LDL-R correlated significantly ( $P \le 0.05$ ) with  $R_f$ , the association was not **as** strong as for other lipoproteins. However, LDL(NCEP) correlated strongly ( $P \le 0.01$ ) probably due to the strong contribution by IDL. The association of HDL<sub>2</sub> with  $R_f$  was the strongest among all measurements. All measurements, except TC, HDLs and Lp[a] cholesterol, were correlated with  $R_f$  in women. Although TC did not correlate significantly, its association approached significance *(P<* 0.1; the corresponding Pvalue for men, however, is **Y0.2);** this is reflected in the association of LDL-R in women only, as LDLR is the major fraction of TC ( $P \le 0.05$ ). In men,  $R_f$  correlated with HDL, HDL<sub>3</sub>, HDL<sub>2</sub>, IDL, and VLDL cholesterol, and TG, but not with LDLR cholesterol, Lp[a] cholesterol, LDL(NCEP) cholesterol, or TC. HDL, HDLs, and

Parameter	All	Women	Men	$P$ (Women vs. Men)
$\mathbf n$	195	68	127	
LDL $Re$	$0.475 \pm 0.071$	$0.51 \pm 0.064$	$0.456 \pm 0.067$	$\leq 0.001$
TC.	$171.6 \pm 30.6$	$174.9 \pm 27.0$	$169.8 \pm 32.3$	$>0.2$ (n.s)
$HDL^{\prime}$	$45.7 \pm 9.8$	$52.4 \pm 9.0$	$42.1 \pm 8.3$	$\leq 0.0001$
HD <sub>1s</sub>	$35.8 \pm 5.6$	$38.4 \pm 5.1$	$34.5 \pm 5.4$	$\leq 0.001$
$HDL_2^b$	$9.8 \pm 6.2$	$14.0 \pm 6.6$	$7.6 \pm 4.7$	$\leq 0.0001$
Lp[a]	$7.7 \pm 4.9$	$7.4 \pm 4.5$	$7.9 \pm 5.2$	$>0.4$ (n.s)
$LDL-Rb$	$88.0 \pm 22.3$	$88.0 \pm 23.4$	$88.0 \pm 21.8$	$>0.9$ (n.s)
$IDL^{\prime}$	$1.5 \pm 5.2$	$10.6 \pm 4.2$	$12.0 \pm 5.6$	< 0.05
LDL(NCEP)*	$107.3 \pm 26.6$	$106.1 \pm 25.7$	$107.9 \pm 27.1$	>0.6(n.s)
<b>VLDL'</b>	$18.5 \pm 9.4$	$16.0 \pm 7.2$	$19.9 \pm 10.1$	50.01
TG	$0.1 \pm 53.8$ (n = 102)	$78.5 \pm 43.8$ (n = 34)	$95.9 \pm 57.6$ (n = 68)	$>0.05$ (n.s)
Age $(y)$	$24.3 \pm 1.6$	$24.4 \pm 1.8$	$24.3 \pm 1.6$	$>0.6$ (n.s)

**TABLE 1. Mean plasma lipoprotein concentrations in medical students selected for this study** 

Values are given as mean ± SD; n.s., nonsignificant (all P values > 0.05 were considered as nonsignificant). Parameter range: TC, 109-269; HDL, 24-78; HDL, 21-50; HDL, 0-36; Lp[a], 0-30; LDL-R, 44-166; IDL, 3-32; LDL(NCEP), 47-198; VLDL, 5-69; TG (mg/dl), 35-393 (only **8 subjects had ,150 mg/dl); and age, 22-29 years. Cholesterol measurements were made using VAP-I method.** 

**%e relative position of the major LDL peak maximum in LDL VAP-I1 profile.** 

**\*Cholesterol concentration (mg/dl).** 

**'LDYNCEP)** = **LDLR** + **Lp[a]** + **IDL.** 



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**Fig.** *6.* Frequency distribution diagram showing the distribution of *f\$* of the major LDL peak maximum in LDL **VAP-I1** density gradient in women and men subjects. *K,* was measured as described in Fig. 2. The mid- $R_t$  position of each cluster is indicated by an arrow.

 $HDL<sub>2</sub>$  cholesterol correlated with  $R<sub>f</sub>$  better in men than women, while IDL correlated better in women. Both LDL(NCEP) and LDL-R cholesterol correlated only in women. The age, either of women or of men, did not significantly correlate with  $R_f$  in this population.

In a forward stepwise multivariate analysis comparing *R,* with sex, VLDL, LDL-R, Lp[a] cholesterol, IDL,  $HDL<sub>3</sub>$ ,  $HDL<sub>2</sub>$ , and triglyceride, only  $HDL<sub>2</sub>$  remained in the model.

To investigate the relationship of LDL subclass distribution with plasma concentrations of various lipoproteins, all subjects were classified into groups based on their major LDL subclass (i.e., the LDL subclass with the highest cholesterol concentration) as obtained from the deconvolution of the parent LDL VAP-I1 profiles. Thus, five distinct groups, with major LDL subclasses LDL-1 to LDL-5, could be identified (LDL-1 and LDL-2 considered as large, buoyant LDL, LDL-3 and LDL-4 as intermediate density LDL, and LDL-5 as small, dense LDL). There were no subjects with LDL-6 as the major subclass in this population. The mean lipoprotein values obtained for each subclass group are summarized in **Table 3.** MDL cholesterol values decreased (the decrease in HDL? being predominant), while IDL and VLDI, cholestcrol, and TG increased as the subclass group density increased; i.e., as subclass group number increased from 1 to 5. LDL-R cholesterol and TC values also increased, but the increase was not continuous. The mean LDL-R values of subclass groups *3,* .4, and 5 (intermediate and small, dense LDL) were similar, while the subclass groups 1 and *2* (large, buoyant LDL) had similar mean values. However, LDL-R mean values of subclass groups *3,* 4, and 5 were higher than the corresponding mean values of subclass groups 1 and 2. The mean LDL-R value for all women subjects in subclass groups 3,4, and 5 together was 94.5 mg/dl compared to the mean value of 81.3 mg/dl for all women subjects in subclass groups 1 and *2* together *(P* < 0.02; data calculated from similar tables prepared, but not shown here, for women and men separately). The corresponding values for men in the two groups (i.e., subclass groups *3,* 4 and 5, and 1 and 2) were 89.8 mg/dl and 82.6 mg/dl, respectively *(P*  > 0.05). Thus, although the differences in mean LDL-R values were observed between the two subgroups within women and men subjects, only the difference within women subgroups was significant. Increases in TC values among subclasses were similar to LDL-R. The data in Table **3** overall suggest that the lipoprotein levels become increasingly atherogenic as LDL subclass group number increases. Thus, mean lipoprotein values for subjects in LDL-5 subclass group are comparatively more atherogenic than the values for subjects in LDL-1 subclass group. Furthermore, 50% of all women were distributed within LDL-1 and LDL-2 subclass groups compared to 25% of all men in the same subclass groups, confirming our observation that compared to men, women generally tend to have more buoyant LDL (Fig. 6) that are accompanied by less atherogenic lipoprotein levels. The progressive changes in the lipoprotein values observed between the subclass groups overall show the effect of LDL subclass distribution on lipoprotein levels.

**Figure 7** shows the frequency distribution diagrams of LDL *Ri* of all five subclass groups plotted separately. With the exception of one subject in the LDL-4 subclass group, the minimal overlap of *Rr* distribution observed between neighboring subclass groups suggests a high accuracy of our mathematical deconvolution program in identifying the major LDL subclass in each subject.

# DISCUSSION

The two most common methods used to study the LDL subspecies distribution have been ultracentrifugation and gradient gel electrophoresis. Currently used ultracentrifugation techniques are lengthy and laborious, and are therefore restricted to research laboratories. Non-denaturing 2-16% gradient gel electrophoresis is a high resolution technique and requires only a small amount of plasma, but the lack of availability of commercial gels could make reproducibility difficult for new users of this technique. **As** the role of LDL subclasses in the development of atherosclerosis becomes

TABLE 2. **Pearson correlation coefficients showing the association of &(relative position of the major** LDL **peak maximum in** LDL VAP-I1 profile; higher the  $R_f$  lower the peak density) with lipoprotein concentration in medical students (n = 195)

		All	Women		Men	
Parameter	r	P	r	P	$\pmb{r}$	$\boldsymbol{P}$
<b>TC</b>	$-0.093$	$>0.1$ (n.s.)	$-0.213$	$<0.1$ (n.s.)	$-0.098$	$>0.2$ (n.s.)
HDL <sup>2</sup>	0.594	50.0001	0.472	< 0.001	0.532	$\leq 0.0001$
HDL <sup>o</sup>	0.350	< 0.001	0.138	$>0.2$ (n.s.)	0.315	0.001
HDL <sub>2</sub> <sup>a</sup>	0.625	< 0.0001	0.532	< 0.001	0.578	< 0.0001
$Lp[a]$ <sup>a</sup>	0.03	$>0.6$ (n.s.)	0.118	$>0.3$ (n.s.)	0.021	$>0.7$ (n.s.)
LDL-R <sup>a</sup>	$-0.152$	< 0.05	$-0.265$	50.05	$-0.108$	$>0.2$ (n.s.)
$IDL^*$	$-0.432$	$\leq 0.0001$	$-0.528$	< 0.001	$-0.379$	$\leq 0.001$
LDL(NCEP) <sup>a</sup>	$-0.206$	50.01	$-0.308$	50.01	$-0.161$	$>0.05$ (n.s.)
VLDL <sup>®</sup>	$-0.386$	50.001	$-0.409$	$\leq 0.001$	$-0.324$	$0.001$
TG	$-0.355$	0.001	$-0.340$	$\leq 0.05$	$-0.325$	50.01
Age	0.009	$>0.9$ (n.s.)	0.105	$>0.3$ (n.s.)	0.066	$>0.4$ (n.s.)

**Weasured as cholesterol concentration (mg/dl); n.s., nonsignificant (all** *P* **values** >0.05 **were considered as nonsignificant).** 

better understood, demand will increase for methods that can rapidly quantify LDL subclasses in a clinical laboratory. The LDL VAP-I1 method provides quantification of cholesterol simultaneously in five LDL subclasses using only 1 h centrifugation time; thus eight samples can be completely analyzed in less than 3 h. In addition, it requires only 70 **pl** of plasma per sample because of its high sensitivity. With 10-fold dilution, plasma samples with **LDLR** cholesterol levels up to 200 mg/dl can be easily analyzed. The method is also highly reproducible, allowing comparisons within and between individuals. Further, this method quantifies cholesterol in LDL density subclasses while the gradient gel electrophoresis method provides sizes of the particles and relative, rather than absolute, concentrations of subspecies. Thus, the two techniques could be complimentary to each other. A good correlation coefficient *(r* = 0.859) observed between  $R_f$  measured by LDL VAP-II and peak LDL particle diameter **as** measured by the gradient gel electrophoresis, an established technique, suggests the reliability of LDL particle distribution measurement by the LDL VAP-I1 technique.

LDL VAP-I1 has the potential to become a useful clinical tool. Subjects can be classified **as** having predominantly small, dense LDL (LDL-5), intermediate density LDL (LDL4 and LDLS), and large, buoyant LDL (LDL-2 and LDL-l), similar to the classification made on the basis of the gradient gel electrophoresis patterns. Only one dense LDL subclass (LDL5) was observed **as** the major subclass in the present population **as** none of the subjects were markedly hypertriglyceridemic (i.e., with TG values >400 mg/dl; only 8 of the 195 subjects had TG >150 mg/dl, with 393 mg/dl being maximum). However, it is possible that more numbers of small, dense LDL subclasses may be observed when markedly hypertriglyceridemic subjects are included.

Although, **as** described above, LDL VAP-I1 compares well with the gradient gel electrophoresis technique, a somewhat greater percentage of subjects were classified as having predominantly intermediate density LDL (LDLS and LDL-4) by LDL VAP-I1 than by the gradient gel electrophoresis **(47%** vs. 34%). There appear to be two possible explanations to this. First, while the gradient gel electrophoresis classification is based on the measurement of size, our classification is based on the flotation rate, a function of both size and density. Although the density of lipoproteins increases with the decrease in size, the relationship between the two physical parameters may not be perfect, i.e., a 1:l relationship may not exist. This is because the variation in 1ipid:protein ratio (which results in density changes) between the subclasses may not result in detectable changes in size or vice versa  $(11)$ . Thus, it is possible that some subjects classified **as** having LDLS **as** their density major subclass (i.e., intermediate density LDL) may indeed have major size LDL subclass with particles larger in diameter than assigned to pattern AB (i.e., pattern A by gradient gel electrophoresis). Second, **as** intermediate density LDL are known to be rich in cholesterol in normal subjects, they more frequently constitute the major subspecies when measured in terms of cholesterol, and this may

**TABLE 3. Comparison of mean lipoprotein concentrations among subjects (n** = **195) classified based on major LDL subclass** 

	<b>LDL</b> Subclass					
Parameter		2	3	4	5	
n	$10 (W = 7, M = 3)$	56 (W = 27, M = 29)	62 (W = 22, M = 40)	51 (W = 11, M = 40)	$16 (W = 1, M = 15)$	
TC	$168.8 \pm 22.1$	$164.2 \pm 24.6$	$175.2 \pm 34.3$	$174.2 \pm 32.6$	$176.7 \pm 31.1$	
HDL <sup>a</sup>	$56.8 \pm 9.0$	$51.9 \pm 10.3$	$44.73 \pm 6.6$	$40.2 \pm 7.7$	$38.2 \pm 6.9$	
HDL <sub>3</sub> <sup>a</sup>	$39.2 \pm 4.5$	$37.5 \pm 5.8$	$36.1 \pm 4.8$	$34.1 \pm 5.6$	$32.8 \pm 6.0$	
HDL <sub>2</sub> <sup>a</sup>	$17.6 \pm 5.2$	$14.4 \pm 7.1$	$8.7 \pm 3.6$	$6.1 \pm 3.8$	$5.4 \pm 3.2$	
$Lp[a]$ <sup>a</sup>	$9.3 \pm 7.5$	$7.4 \pm 5.1$	$7.3 \pm 3.8$	$8.9 \pm 5.5$	$5.7 \pm 3.6$	
LDL-R <sup>a</sup>	$81.3 \pm 24.1$	$81.7 \pm 19.1$	$91.4 \pm 25.7$	$90.8 \pm 20.8$	$92.4 \pm 19.3$	
$IDL^c$	$6.9 \pm 3.0$	$8.6 \pm 3.7$	$12.6 \pm 5.2$	$13.3 \pm 5.1$	$14.6 \pm 4.8$	
LDL(NCEP) <sup>a</sup>	$97.5 \pm 23.9$	$97.7 \pm 21.3$	$111.3 \pm 30.5$	$113.0 \pm 25.8$	$112.7 \pm 22.5$	
VLDL <sup>a</sup>	$11.4 \pm 3.9$	$14.7 \pm 7.4$	$19.1 \pm 9.6$	$21.0 \pm 8.1$	$26.2 \pm 12.6$	
<b>TG</b>	$54.0 \pm 12.2$	$73.3 \pm 33.0$	$91.9 \pm 67.3$	$103.7 \pm 44.5$	$185.3 \pm 55.3$	
Age(y)	$24.7 \pm 1.9$	$24.5 \pm 1.6$	$24.1 \pm 1.5$	$24.4 \pm 1.6$	$24.6 \pm 2.1$	

**Major LDL subclass is the LDL subcurve with the highest area as obtained from deconvolution program; W, women; M, men. Subjects were medical students selected for this study.** 

**"Cholesterol concentration (mg/dl).** 

not be true when measured in terms of other properties, such as size. However, the percentages of prevalence of intermediate density LDL obtained in the present study by both the techniques were much higher than the corresponding reported value of 15% in normolipidemic subjects (30).

As observed in other studies, the gender differences in the distribution of LDL subclasses were also evident in the present study (Table 3). Small, dense LDL was substantially less prevalent in women subjects compared to men subjects. Only 1 of the 68 (1.5%) women subjects studied here showed LDL-5 (small, dense LDL) **as** the major LDL subclass compared to 15 of the 127 (12%) men subjects. Similarly, fewer number of women subjects ( $n = 11$ , 16%) had LDL-4 (an intermediate density LDL subclass) compared to men subjects ( $n = 40, 31.5\%$ ). However, the prevalence of LDL3 (also an intermediate density LDL subclass) was similar in both women and men subjects (32.5% and 31.5%, respectively). On the other hand, a greater percentage of women subjects had LDLl and LDL2 (large, buoyant LDL) as their major LDL subclass compared to men subjects (40% and 10% of LDL2 and LDLl, respectively, in women compared to 23% and 2.5%, respectively, in men). Thus, women subjects had increased prevalence of large, buoyant LDL compared to men subjects, and both groups had significantly increased prevalence of intermediate density LDL.

In the present work, the relative position of the major LDL peak maximum in the density gradient  $(R_f; a$  higher *Rf* indicates more buoyant LDL) was found to associate

the results obtained from other investigators (5, **6,** 22, 23, 31, 32), some notable differences were observed which appear to be due to the specific age range of this population (22-29 years). The association of TG with *Rf*  was not **as** strong **as** found in other studies. This appears to be primarily due to the lower TG values in this population; the mean TG value being only 90 mg/dl, a value smaller than the cross-over value (95 mg/dl) between patterns A and B, suggested by Austin et al. (12). On the other hand, HDL and HDL2 cholesterol showed strong association with  $R_f$  especially in men. Although association of HDLs was not **as** strong **as** HDL or HDL2 in men, it was highly significant ( $P \le 0.001$ ). However,  $HDL<sub>3</sub>$  did not correlate with  $R<sub>f</sub>$  in women. The age, within the selected narrow and lower range (22-29 years), was not correlated with  $R_f$  in both women and men subjects. The association of Lp[a] cholesterol with LDL size or density has not been previously studied in detail. The only report available in the literature suggests a significant positive association of Lp[a] **mass** with dense LDL in women only (33). However, our data suggested no such association, at least in the present population. The overall agreement of our results with

strongly and positively with HDL, HDLs, and HDL2 cholesterol, and negatively with VLDL and IDL cholesterol and TG. LDL(NCEP) cholesterol, which was derived by adding Lp[a] and IDL cholesterol to LDLR, correlated better than LDL-R with  $R_f$  in women. This increased association is due most likely to the contribution by IDL, which also correlated better in women than in men. While these results are generally consistent with

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**Rf (Relative position of the major LDL peak in the gradient)** 

**D.64** 

 $\ddot{9}$  $58$ 

 $58$ 

*Fig.* **7.** Frequency distribution **diagram** showing the distribution of &of the major **LDL** peak maximum in **LDL VAP-XI** profiles of subjects classified based on their major **LDL subclass.** All subjects (n = **195)**  were classified into five group **(LDLI** through **LDL5)** based on their major **LDL subclass as** determined by the mathematical deconvolution software.  $R_f$  of the major LDL peak maximum was determined as described in Materials and Methods and the frequency distribution diagram of *Rj* for each group **was** plotted. Minimal overlap of the *R/*  distribution between neighboring groups suggests a high accuracy of the mathematical deconvolution in the identification of major **LDL subclass.** 

those previously reported again suggests the reliability of the LDL VAP-11.

Another important feature of LDL VAP-I1 is its ability **to** classify subjects based on the atherogenecity of various lipoprotein values. For example, subjects classified in LDL5 subclass group had all lipoproteins at more atherogenic levels **than** the lipoprotein levels in subjects in LDLl subclass group (Table 3). In addition, the

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ant LDL su progressive changes in atherogenecity of all lipoprotein levels, except LDLR, can be seen from one to the next subgroup. The changes in LDL-R and LDL (NCEP) were not continuous, **as also** observed by Campos et **al.** (15). The mean LDLR and LDL(NCEP) cholesterol values of both men and women subjects in LDL-3 and LDL-4 subclass groups (intermediate density LDL subclass) were higher than the corresponding mean values **ob**served for subjects with LDL-1 and LDL-2 (large, buoyant LDL subclass) **as** their major LDL subclass (data not shown). These observations are similar to the observations made by Campos et al. (15). However, only the difference between LDL-R mean values of LDL2 and LDLS subclass groups in women and the differences between mean values of LDL(NCEP) of LDL-2 and LDLS subclass groups in women and LDL2 and LDL-4 subclass groups in men were significant  $(P \le 0.05)$ . In addition, in our study the mean LDL-R and LDL(NCEP) values in subjects (both men and women) with LDL-5 (small, dense LDL) **as** the major subclass were similar to the corresponding values in subjects with LDL-4 and LDLS as their major LDL subclass, while LDL cholesterol values decreased for subjects with small, dense LDL (except in men with LDL5) in the study by Campos et **al.** (15).

> Thus, our results overall suggest that LDL VAP-I1 can also serve **as** a screen for atherogenic lipoproteins. In summary, we have developed a simple and rapid method for the determination of LDL subclasses that can be utilized for clinical studies. The LDL VAP-I1 method can be used to study the LDL subclass cholesterol distribution in small animals and infants where the blood volume obtained is limited.

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